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Please find below and/or attached an Office communication concerning this application or proceeding.

The time period for reply, if any, is set in the attached communication.

Office Action Summary

Application No.

10/658,355

Applicant(s)

GANTIER ET AL.

Examiner

Russell S. Negin

Art Unit

1631

-- The MAILING DATE of this communication appears on the cover sheet with the correspondence address --

Period for Reply

A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE 3 MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION.

- Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication.
- If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication.
- Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b).

Status

- 1) ☒ Responsive to communication(s) filed on 04 September 2007.
- 2a) ☐ This action is **FINAL**. 2b) ☒ This action is non-final.
- 3) ☐ Since this application is in condition for allowance except for formal matters, prosecution as to the merits is closed in accordance with the practice under *Ex parte Quayle*, 1935 C.D. 11, 453 O.G. 213.

Disposition of Claims

- 4) ☒ Claim(s) 1-7, 9-28, 30-50, 53-67 and 79-89 is/are pending in the application.
- 4a) Of the above claim(s) 12-14, 19-27, 39-50, 56-67 and 82-89 is/are withdrawn from consideration.
- 5) ☐ Claim(s) _____ is/are allowed.
- 6) ☒ Claim(s) 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 is/are rejected.
- 7) ☒ Claim(s) 28 is/are objected to.
- 8) ☐ Claim(s) _____ are subject to restriction and/or election requirement.

Application Papers

- 9) ☐ The specification is objected to by the Examiner.
- 10) ☐ The drawing(s) filed on _____ is/are: a) ☐ accepted or b) ☐ objected to by the Examiner.
Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a).
Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d).
- 11) ☐ The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152.

Priority under 35 U.S.C. § 119

- 12) ☐ Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f).
- a) ☐ All b) ☐ Some * c) ☐ None of:
1. ☐ Certified copies of the priority documents have been received.
 2. ☐ Certified copies of the priority documents have been received in Application No. _____.
 3. ☐ Copies of the certified copies of the priority documents have been received in this National Stage application from the International Bureau (PCT Rule 17.2(a)).

* See the attached detailed Office action for a list of the certified copies not received.

Attachment(s)

- | | |
|--|---|
| 1) <input checked="" type="checkbox"/> Notice of References Cited (PTO-892) | 4) <input type="checkbox"/> Interview Summary (PTO-413)
Paper No(s)/Mail Date. _____ |
| 2) <input type="checkbox"/> Notice of Draftsperson's Patent Drawing Review (PTO-948) | 5) <input type="checkbox"/> Notice of Informal Patent Application |
| 3) <input checked="" type="checkbox"/> Information Disclosure Statement(s) (PTO/SB/08)
Paper No(s)/Mail Date <u>9/4/07; 9/17/07</u> . | 6) <input type="checkbox"/> Other: _____ |

DETAILED ACTION

Comments

Applicants' amendments and request for reconsideration in the communication filed on 4 September 2007 are acknowledged and the amendments are entered.

Claims 1-7, 9-28, 30-50, 53-67, and 79-89 are pending and claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 are examined in this Office action.

Information Disclosure Statement

The information disclosure statements filed on 4 September 2007 and 17 September 2007 have been considered in full.

Withdrawn Objections/Rejections

The objections to claims 1, 52, and 55 because of informalities are withdrawn in view of amendments to the set of claims filed on 4 September 2007.

The rejections of claims 1-11, 15-18, 28-38, 51-55, and 79-81 under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention are withdrawn in view of amendments to the set of claims filed on 4 September 2007.

The rejections of claims 1, 4-11, 28-35, 51-52, and 80 under 35 U.S.C. 103(a) as being unpatentable over Ladner et al. [USPAT 5,096,815] in view of Stabach et al.

[Biochemistry, 1997, volume 26, pages 57-65] are withdrawn in view of arguments filed by applicant on pages 24-26 of the Remarks.

The rejections of claims 1, 6, 15-18, 28, 36-38, 52-55, and 81 under 35 U.S.C. 103(a) as being unpatentable over Ladner et al. in view of Stabach as applied to claims 1, 4-11, 28-35, 51-52, and 80 above, in further view of Alam et al. [Journal of Biotechnology, volume 65, 1998, pages 183-190] are withdrawn in view of arguments filed by applicant on pages 24-26 of the Remarks.

The rejections of claims 1-3 under 35 U.S.C. 103(a) as being unpatentable over Ladner et al. in view of Stabach as applied to claims 1, 4-11, 28-35, 51-52, and 80 above, in further view of Chiang et al. [Annual Reviews of Microbiology, 1999, volume 53, pages 129-154] are withdrawn in view of arguments filed by applicant on pages 24-26 of the Remarks.

The rejections of claims 1 and 79 under 35 U.S.C. 103(a) as being unpatentable over Ladner et al. in view of Stabach as applied to claims 1, 4-11, 28-35, 51-52, and 80 above, in further view of Jones et al. [CABIOS, volume 8, 1992, pages 275-282] are withdrawn in view of arguments filed by applicant on pages 24-26 of the Remarks.

Claim Rejections - 35 USC § 112

The following is a quotation of the second paragraph of 35 U.S.C. 112:

The specification shall conclude with one or more claims particularly pointing out and distinctly claiming the subject matter which the applicant regards as his invention.

Claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 are rejected under 35 U.S.C. 112, second paragraph, as being indefinite for failing to particularly point out and distinctly claim the subject matter which applicant regards as the invention.

Claim 1 recites the limitation "the evolved predetermined property or activity" in line 10. There is insufficient antecedent basis for this limitation in the claim. Although the claim recites a "predetermined property or activity," the claim does not recite an "evolved predetermined property or activity."

Claim 1, line 16 and claim 28, line 14 each recite "a restricted subset," where it is unclear as to whether this restricted subset is the same restricted subset as that recited in line 15 of claim 1 and lines 12-13 of claim 28, or a different restricted subset.

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(e) the invention was described in (1) an application for patent, published under section 122(b), by another filed in the United States before the invention by the applicant for patent or (2) a patent granted on an application for patent by another filed in the United States before the invention by the applicant for patent, except that an international application filed under the treaty defined in section 351(a) shall have the effects for purposes of this subsection of an application filed in the United States only if the international application designated the United States and was published under Article 21(2) of such treaty in the English language.

Claims 28 and 30-35 are rejected under 35 U.S.C. 102(e) as being anticipated by Winter [US Patent 6,548,640; issued 15 April 2003; filed 26 May 1995].

Claims 28 and 30-35 are drawn to a method for generating proteins with a desired property or activity produced by generating mutants of a protein, and inserting into host cells nucleic acids which encode for the mutant proteins.

The invention of Winter studies altered antibodies that have a heavy or light chain variable domain in which the framework regions differ from the framework regions naturally associated with the complementarity determining regions of the variable domain and in which the framework regions are derived from the source of framework regions that differs from the framework regions naturally associated with the complementarity determining regions of the variable regions.

Claim 28 is drawn to a method for generating proteins with a desired property or activity. As described in the abstract and Table 3 in column 20 of Winter, antibodies are studied and altered to assess activity towards antigens.

The first step of the instant claim is identifying residues in a target protein *in silico* that are associated with the property, and designating the loci of such residues is-HIT loci.

Column 21, lines 13-30 of Winter teach the use of an *in silico* generated representation of the loop of Phe27 to Tyr35 in the heavy chain variable domain of the human myeloma protein KOL which is crystallographically solved. From this *in silico* generation of the loop, Ser27 is selected to be mutated to Phe and Ser30 is selected to be mutated to Thr.

The second step of the instant method is preparing variant nucleic acid molecules encoding variant proteins, wherein each variant nucleic acid encodes a

candidate LEAD mutant protein that differs by one replacement amino acid at one is-HIT locus from the target protein wherein: the amino acid residues at each of the identified is-HIT target loci in the target protein is replaced with all of the non-native amino acids, or the amino acid residues at each of the identified is-HIT target loci in the target is replaced with a restricted subset of the remaining 19 non-native amino acids; and a restricted subset is a group of selected amino acids selected to have a predetermined effect on protein activity.

The third step of the instant method is separately introducing the nucleic acid molecules encoding each candidate LEAD protein into hosts for expression thereof, and expressing the nucleic acid molecules encoding each variant protein to produce sets of candidate LEAD proteins wherein: each candidate LEAD protein in a set contains the same amino acid replacement; each candidate LEAD protein contains a single amino acid replacement, and differs from the target protein by one amino acid replacement.

The second and third steps of the instantly rejected claims are taught in Winter in column 19, line 62 to column 20, line 7, which states:

In stage 1, the pSVgpt vectors HuVHCAMP-RalgG2B, and also two mutants for reasons to be explained below, HuVHCAMP(Ser27 to Phe)-RalgG2B, HuVHCAMP(Ser27 to Phe, Ser30 to Thr)-RalgG2B) [sic] were introduced into the heavy chain loss variant of YTH34.5HL. In stage 2, the pSVgpt vectors RaVHCAMP-RalgG2B, RaCVHCAMP-HulgG1, RaVHCAMP-HulgG2, RaVHCAMP-HulgG3, RaVHCAMP-HulgG4 were transfected as described above. In stage 3, the pSV-gpt vector Hu(Ser27-Phe, Ser30-Thr)VHCAMP-HulgG1 was cotransfected with the pSV-neo vector HuVLCAMP-HulgK into the rat myeloma cell line Y0 (Y B2/3.0 Ag 20).

This passage in Winter describes the cell line used to produce the nucleic acids that transcribe into the wild type and mutant proteins taught in column 21, lines 13-30 of Winter. Each vector type produces the unique type of protein (either the original or

mutant). The amino acid serine is replaced with the amino acid phenylalanine, thus phenylalanine constitutes a “restricted subset” of amino acids used to replace the original amino acid in the unmodified protein. All of the single mutant proteins contain the same Ser27 to phenylalanine replacement.

The fourth step of the instant method individually screens each set of variant LEAD candidate proteins to identify any that have an activity or property that differs by a predetermined amount from the activity of the unmodified target protein, thereby identifying proteins that are LEADs.

Table 3 of Winter in column 20 lists the results of screening the respective antibody proteins for antigen binding. Specifically, Table 3 illustrates the concentrations of antibody in ug/ml at 50% binding or lysis. In the original protein, concentration of 27.3 ug/ml is required for 50% binding while in the mutant proteins (Ser27 to Phe), 1.8 ug/ml is required for 50% binding. Consequently, the mutant LEAD protein has a much higher affinity for antigen as the original protein. Column 14, lines 52-59 and column 20, line 66 to column 22, line 12 indicate a predetermined change of binding to antigen upon mutating such residues in the “hypervariable region” including Ser27 and Ser30.

Claim 30 is further limiting wherein each of the residues at identified is-HIT loci in the target protein is replaced with codons encoding a restricted subset of the remaining 19 amino acids, wherein a restricted subset is a group of selected amino acids selected to have a predetermined effect on protein activity.

As recited in Winter in column 19, line 62 to column 20, line 7, codons are replaced in nucleic acids to encode the proteins listed in Table 3 of Winter. In this instance, a serine is replaced by a phenylalanine.

Claim 31 is further limiting wherein the total number of is-HIT loci which are replaced with replacement amino acids is less than the total number of amino acid residues within the full-length of the target protein.

In Winter, a single loci, amino acid number 27, is replaced. The protein has more than a single residue, so this substitution meets the limitations of the instant claim 31.

Claim 32 is further limiting wherein each of the residues at identified is-HIT loci in the target protein is replaced with a restricted subset of the remaining 19 amino acids; and the total number of is-HIT loci that is replaced with replacement amino acids is less than the total number of amino acid residues within the full-length of the target protein.

As taught by Winter in column 19, line 62 to column 20, line 7, codons are replaced in nucleic acids to encode the proteins listed in Table 3 of Winter. In this instance, a serine is replaced by a phenylalanine. This phenylalanine constitutes the restricted subset of the 19 remained amino acids.

In Winter, a single loci, amino acid number 27, is replaced from a serine to a single type of residue, a phenylalanine. The protein has more than a single residue, so this substitution fits the teachings of the instant claim 31.

Claim 33 is further limiting with three additional steps comprising:

- generating a population of sets of nucleic acid molecules encoding sets of candidate super-LEAD proteins wherein;

- each candidate super-LEAD protein comprises a combination of two or more of the single amino acid mutations derived from two or more LEAD mutant proteins; and each set encodes a single candidate super-LEAD protein;

- introducing each set of nucleic acid molecules encoding candidate super-LEADs into cells and expressing the encoded candidate super-LEAD proteins;

From the discussion above, one of the mutant proteins fits the description of a candidate super-LEAD protein. Specifically, the double mutant (Ser27 to Phe, Ser30 to Thr) has two mutations from the original antibody protein. This double mutant is generated from cells with vectors encoding for this double mutant.

- individually screening the sets of encoded candidate super-LEAD proteins to identify one or more proteins that has activity that differs from the unmodified target protein and has properties that differ from the unmodified target protein and has properties that differ from the original LEADs, wherein each such protein is designated a super-LEAD.

Table 3 of column 20 of Winter tabulates the antigen screening of such a double mutant. Specifically, Table 3 illustrates the concentrations of antibody in ug/ml at 50% binding or lysis. In the original protein, concentration of 27.3 ug/ml is required for 50%

binding while in the mutant proteins (Ser27 to Phe), 1.8 ug/ml is required for 50% binding. The double mutant protein (Ser27 to Phe and Ser 30 to Phe) requires a concentration of 2.0 ug/ml for 50% binding. Consequently, the mutant LEAD protein and super-LEAD proteins have a much higher affinity for antigen as the original protein.

Claim 34 is further limiting wherein one of the types of mutations is site specific mutagenesis.

As illustrated in Table 3 in column 20 of Winter, the nucleic acids are mutated at sites corresponding to residues 27 and 30.

Claim 35 is further limiting wherein there are two positions generated from a single nucleic acid molecule. In the instance of Winter, the amino acid positions are amino acids 27 and 30, which are mutated for desired activities towards antigens.

Claim Rejections - 35 USC § 103

The following is a quotation of 35 U.S.C. 103(a) which forms the basis for all obviousness rejections set forth in this Office action:

(a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negated by the manner in which the invention was made.

This application currently names joint inventors. In considering patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein

were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).

The following rejections are newly applied:

35 U.S.C. 103 Rejection #1:

Claims 1-7, 9-11, and 15 are rejected under 35 U.S.C. 103(a) as being unpatentable over Winter as applied to claims 28, and 30-35 above, in further view of Chiang et al. [Annual Reviews in Microbiology, 1999, volume 53, pages 129-154].

Claim 1 is drawn to the same subject matter as instant claim 28, with the additional limitation of locating each of the host cells with nucleic acids encoding for the mutant proteins into a different locus on the array.

The invention of Winter teaches a method of generating a protein having a predetermined property, but does not teach use of an array.

The study of Chiang et al. investigates the *in vivo* genetic analysis of bacterial virulence. Specifically, one of the techniques described in Chiang et al. is the "STM" strategy diagrammed in Figures 4 and 5 of Chiang et al. on pages 140-141. In this strategy, arrays are formed where each cell expresses nucleic acids that encode specific proteins. The use of arrays by Chiang et al. allows for more systematic analysis and comparison of the mutants for the purpose in Chiang et al. of *in vivo* analysis.

Claim 2 is further limiting wherein the array comprises a solid support with separate loci and each set of cells is at a different locus.

Claim 3 is further limiting wherein the loci comprise wells; and each well contains one set of cells.

Figure 5 of Chiang et al. illustrates such microarrays with such loci comprising individual wells.

Claim 4 is further limiting wherein the nucleic acid molecules comprise plasmids, and the cells are eukaryotic cells that are transfected with the plasmids or are bacterial cells are transformed with the plasmids.

Claim 5 is further limiting wherein the nucleic acid molecules are produced by site specific mutagenesis.

The passages cited from Winter indicate that the cells that produce the antibodies are lymphoid cells transfected with plasmids with the vectors described in column 19, lines 32-47.

The site specific mutagenesis in Winter is at the codon corresponding to residue number 27 of the protein.

Claims 6 and 7 recite the same limitations as claims 33 and 34 with the exception of being dependent from two different base claims. The limitations are disclosed in

Winter, as described in the 35 U.S.C. 102(e) rejection above. The array limitations are disclosed in Chiang et al.

Claim 9 is further limiting wherein the replacement amino acids correspond to the 19 remaining amino acids.

Claim 10 is further limiting wherein the nucleic acids are produced systematically by replacing each codon that is an is-HIT with one or more codons encoding a restricted subset of the remaining amino acids.

As recited in Winter in column 19, line 62 to column 20, line 7, codons are replaced in nucleic acids to encode for the proteins listed in Table 3 of Winter. In this instance, a serine is replaced by a phenylalanine.

Claim 11 is further limiting wherein the number of LEAD amino acid positions constitute two amino acid positions.

As described above, in Winter, two amino acid positions are replaced (positions 27 and 30).

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the mutagenesis study of Winter by use of the array methods of Chiang et al. where the motivation would have been that the array methods of Chiang et al. allow for a more systematic means for comparing and analyzing different mutants (see, for example, pages 140-141 of Chiang et al.)

35 U.S.C. 103 Rejection #2:

Claims 36-38 are rejected under 35 U.S.C. 103(a) as being unpatentable over Winter as applied to claims 1-7, 9-11, 15, 28, and 30-35 above, and further in view of Alam et al. [Journal of Biotechnology, volume 65, 1998, pages 183-190].

Claim 36 is further limiting wherein each is-HIT target residue is susceptible to digestion by one or more proteases.

Claim 37 is further limiting wherein the LEADs or super-LEADs possess increased resistance to proteolysis compared to unmodified target protein.

Claim 38 is further limiting wherein each is-HIT target residue is resistant to digestion by one or more proteases.

The invention of Winter teaches the method of generating a protein having a predetermined property, as set forth above.

Winter does not teach use of proteolysis as a means of digestion.

In the article of Alam et al., entitled, "Expression and purification of a mutant human growth hormone that is resistant to proteolytic cleavage by thrombin, plasmin and human plasma in vitro," Alam et al. take a section of human growth hormone which is not resistant to proteolysis, and conduct mutations to the hormone to make it resistant to proteolysis (see for instant, abstract on page 183 which states, "In this study, oligonucleotide primer-directed mutagenesis was used to produce recombinant mutant hGHs resistant to limited proteolysis by these proteases.")

Susceptibility and resistance to proteolysis are determined by whether the mutation is conducted or whether the wild type is sustained, respectively.

Alam et al. ends their article in column 2 of page 189 by stating:

This mutant GH [growth hormone] modified at the proteolytically sensitive sites is expected to have a longer period of bioavailability with characteristic pharmacological importance, implicating a potent clinical application in the future. In that case, however, the possibility that the mutant hGH may raise the antibody during the treatment has to be investigated in detail.

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the is-Hit generation method of Winter by generating proteolysis resistance mutants, as described in Alam et al. where the motivation would have to generate proteins which are expected to have a longer period of bioavailability with characteristic pharmacological importance [see column 2, page 189 of Alam et al.]

35 U.S.C. 103 Rejection #3:

Claims 16-18, 53-55, and 81 are rejected under 35 U.S.C. 103(a) as being unpatentable over Winter in view of Chiang et al. as applied to claims 1-7, 9-11, 15, 28, and 30-35 above, and further in view of Alam et al. [Journal of Biotechnology, volume 65, 1998, pages 183-190].

Claim 16 is further limiting wherein each is-HIT target residue is susceptible to digestion by one or more proteases.

Claim 17 is further limiting wherein the LEADs or super-LEADs possess increased resistance to proteolysis compared to unmodified target protein.

Claim 18 is further limiting wherein each is-HIT target residue is resistant to digestion by one or more proteases.

Claim 53 is further limiting wherein a predetermined property is susceptibility to digestion by proteases.

Claim 54 is further limiting wherein the LEADs possess increased resistance to proteolysis compared to unmodified target protein.

Claim 55 is further limiting wherein the predetermined property is resistance to digestion by one or more proteases.

Claim 81 is further limiting wherein the LEADs possess increased resistant to proteolysis compared to the original protein.

Winter and Chiang et al. make obvious the ability to generate LEAD proteins in arrays, as set forth above.

Winter and Chiang et al. do not teach use of proteolysis as a means of digestion.

In the article of Alam et al., entitled, "Expression and purification of a mutant human growth hormone that it resistant to proteolytic cleavage by thrombin, plasmin and human plasma in vitro," Alam et al. take a section of human growth hormone which is not resistant to proteolysis, and conduct mutations to the hormone to make it resistant to proteolysis (see for instant, abstract on page 183 which states, "In this study, oligonucleotide primer-directed mutagenesis was used to produce recombinant mutant hGHs resistant to limited proteolysis by these proteases.")

Susceptibility and resistance to proteolysis are determined by whether the mutation is conducted or whether the wild type is sustained, respectively.

Alam et al. ends their article in column 2 of page 189 by stating:

This mutant GH [growth hormone] modified at the proteolytically sensitive sites is expected to have a longer period of bioavailability with characteristic pharmacological importance, implicating

a potent clinical application in the future. In that case, however, the possibility that the mutant hGH may raise the antibody during the treatment has to be investigated in detail.

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the is-Hit generation method of Winter and Chiang et al. by generating proteolysis resistance mutants, as described in Alam et al. where the motivation would have been to generate proteins which are expected to have a longer period of bioavailability with characteristic pharmacological importance [see column 2, page 189 of Alam et al.]

35 U.S.C. 103 Rejection #4:

Claim 79 is rejected under 35 U.S.C. 103(a) as being unpatentable over Winter in view of Chiang et al. as applied to claims 1-7, 9-11, 15, 28, and 30-35 above, and further in view of Jones et al. [CABIOS, volume 8, 1992, pages 275-282].

Claim 79 is further limiting wherein the replacement amino acids are selected using Percent Accepted Mutations (PAM) matrices.

Winter and Chiang et al. make obvious the ability to generate LEAD proteins in arrays, as set forth above.

Winter and Chiang et al. do not teach PAMs.

The study of Jones et al., entitled "The rapid generation of mutation data matrices from protein sequences," shows PAM matrices on Table I on page 279 for the purpose of mutation of protein sequences.

Jones et al. explain the purpose of using these specific techniques at the end of the third full paragraph of column 1 of page 276:

... it is our hope that the matrices presented here will more clearly express the general nature of the underlying amino acid similarities.

It would have been obvious to someone of ordinary skill in the art at the time of the instant invention to modify the is-HIT generation method of Winter and Chiang et al. by further use the PAMs described in Jones et al. where the motivation would have been that applying the site directed mutagenesis study to the claimed analysis condition of PAM matrices yields a clearer and more efficient understanding of the amino acid residues comprising the protein of interest (see for example, page 276 of Jones et al.)

Response to Arguments

Applicant's arguments and declaration with respect to claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 have been considered but their arguments are moot in view of the new ground(s) of rejection.

Although the references of Chiang et al., Alam et al., and Jones et al. are sustained from the previous Office actions, applicants' arguments are specifically related to their motivations of combinations with the previously used reference of Ladner et al. and Stabach et al. Since Ladner et al. and Stabach et al. are not utilized in this rejection, these arguments are moot in view of the new grounds of rejection.

Double Patenting

The nonstatutory double patenting rejection is based on a judicially created doctrine grounded in public policy (a policy reflected in the statute) so as to prevent the unjustified or improper timewise extension of the "right to exclude" granted by a patent

and to prevent possible harassment by multiple assignees. A nonstatutory obviousness-type double patenting rejection is appropriate where the conflicting claims are not identical, but at least one examined application claim is not patentably distinct from the reference claim(s) because the examined application claim is either anticipated by, or would have been obvious over, the reference claim(s). See, e.g., *In re Berg*, 140 F.3d 1428, 46 USPQ2d 1226 (Fed. Cir. 1998); *In re Goodman*, 11 F.3d 1046, 29 USPQ2d 2010 (Fed. Cir. 1993); *In re Longi*, 759 F.2d 887, 225 USPQ 645 (Fed. Cir. 1985); *In re Van Ornum*, 686 F.2d 937, 214 USPQ 761 (CCPA 1982); *In re Vogel*, 422 F.2d 438, 164 USPQ 619 (CCPA 1970); and *In re Thorington*, 418 F.2d 528, 163 USPQ 644 (CCPA 1969).

A timely filed terminal disclaimer in compliance with 37 CFR 1.321(c) or 1.321(d) may be used to overcome an actual or provisional rejection based on a nonstatutory double patenting ground provided the conflicting application or patent either is shown to be commonly owned with this application, or claims an invention made as a result of activities undertaken within the scope of a joint research agreement.

Effective January 1, 1994, a registered attorney or agent of record may sign a terminal disclaimer. A terminal disclaimer signed by the assignee must fully comply with 37 CFR 3.73(b).

Claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 are provisionally rejected on the ground of nonstatutory obviousness-type double patenting as being unpatentable over claims 1-3, 6-16, and 19 of copending Application No. 11/707,014. Although the conflicting claims are not identical, they are not patentably distinct from each other

because the claims of '014 are a species of the generic claims listed in the instant application, and therefore the claims of '014 anticipate the claims of the instant application. While both sets of claims are drawn to identifying LEAD or super-LEAD proteins that possess a desired activity, '014 has the additional limitation of choosing the is-HITs based on structural parameters.

This is a provisional obviousness-type double patenting rejection because the conflicting claims have not in fact been patented.

Claim Rejections - 35 USC § 102

The following is a quotation of the appropriate paragraphs of 35 U.S.C. 102 that form the basis for the rejections under this section made in this Office action:

A person shall be entitled to a patent unless –

(f) he did not himself invent the subject matter sought to be patented.

Claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81 are rejected under 35 U.S.C. 102(f) because the applicant did not invent the claimed subject matter.

For the reasons discussed above, it is apparent that copending Application No. 11/707,014 contains claimed subject matter in claims that is not patentably distinct from instant claims 1-7, 9-11, 15-18, 28, 30-38, 53-55, 79 and 81. Because the inventive entity of copending Application 11/707,014 is different from the instant application, a rejection is appropriate under 35 U.S.C. 102(f). This rejection could be overcome by amendment of the appropriate claims so that the claims are patentably distinct, or by

filing a declaration stating the inventive entity for the commonly claimed subject matter is identical.

Conclusion

No claim is allowed.

Papers related to this application may be submitted to Technical Center 1600 by facsimile transmission. Papers should be faxed to Technical Center 1600 via the central PTO Fax Center. The faxing of such pages must conform with the notices published in the Official Gazette, 1096 OG 30 (November 15, 1988), 1156 OG 61 (November 16, 1993), and 1157 OG 94 (December 28, 1993)(See 37 CFR § 1.6(d)). The Central PTO Fax Center Number is (571) 273-8300.

Any inquiry concerning this communication or earlier communications from the examiner should be directed to Russell Negin, Ph.D., whose telephone number is (571) 272-1083. The examiner can normally be reached on Monday-Friday from 7am to 4pm.

If attempts to reach the examiner by telephone are unsuccessful, the examiner's Supervisor, Marjorie Moran, Supervisory Patent Examiner, can be reached at (571) 272-0720.

Information regarding the status of the application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only.

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/Marjorie A. Moran/
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12/5/2007